NUS/T1p-NMR techniques for studying receptor-ligand interactions on living cells surface

Clementina Acconcia¹, Biancamaria Farina², Maria Teresa Gentile¹, Sonia Di Gaetano³, Domenica Capasso⁴, Annarita Del Gatto³, Antonella Paladino⁵, Michele Saviano⁶, Roberto Fattorusso¹, Laura Zaccaro³ and Luigi Russo¹.

¹Department of Environmental, Biological and Pharmaceutical Science and Technology, University of Campania -Luigi Vanvitelli, via Vivaldi 43, 81100 Caserta, Italy.

²Advanced Accelerator Applications, a Novartis Company, via Vivaldi 43, 81100 Caserta, Italy.

³Institute of Biostructures and Bioimaging-CNR, via Mezzocannone 16, 80134, Naples, Italy.

⁴Interdepartmental Center of Bioactive Peptide, University of Naples Federico II, Via Mezzocannone 16, 80134 Naples, Italy.

⁵Department of Science and Technology, University of Sannio, via Francesco de Sanctis, Benevento 82100, Italy. ⁶Institute of Crystallography-CNR, via Amendola 122/O, 70126 Bari, Italy.

*e-mail: clementina.acconcia@unicampania.it

Nuclear Magnetic Resonance Spectroscopy represents a powerful technique for studying, at atomic resolution, protein-protein and protein-ligand interactions, directly in the intracellular environment or on the membrane surface of living cells. In- and on- cell NMR methods for observations of ligands, including transfer NOESY (trNOESY) and saturation transfer difference (STD). However, the application of both NMR methodologie is limited by the short lifetime of cells in an NMR tube, which often prevents the acquisition of experiments longer than few hours [1,2]. To overcome these limitations, we developed an on-cell NMR-based strategy for describing the molecular determinants driving the receptor-ligand recognition process under native conditions. This approach relies on the combination of high-resolution NMR data with Molecular Dynamics simulations and Molecular Docking studies. The key point of our strategy is the application of Non-Uniform Sampling (NUS) and T1p-NMR techniques to collect atomicresolution structural and dynamics information on the receptor-ligand interactions using living cells. We tested our approach to characterize the recognition mechanism of the $\alpha_{v}\beta_{5}$ -integrin by RGDechi15D peptide [3,4]. Our data demonstrate that the developed strategy represents an alternative on-cell NMR tool for studying, at atomic resolution, receptor-ligand recognition mechanism on living cells surface.

References

^[1] E. Luchinat, L. Banci IUCrJ. 2017, 4, 108-18.

^[2] E. Luchinat, L. Banci J Biol Chem. 2016, 291, 3776-84.

^[3] B. Farina, I.de Paola, L. Russo, D. Capasso, A. Liguoro, A. Del Gatto, et al. Chemistry 2016, 22, 681-93.

^[4] D. Capasso, A. Del Gatto, D. Comegna, L. Russo, R. Fattorusso, M. Saviano, et al. Molecules 2020, 25.